

DIAGNOSTIC METHODS FOR DRUG SCREENING FOR ALZHEIMER'S DISEASE

Technical Field

The field of this invention is the screening of drugs for treatment of Alzheimer's and related neurodegenerative diseases.

5 Background

Alzheimer's disease (AD) is the most common cause of dementia in the elderly. Mutations in the amyloid precursor protein gene (*APP*) and presenilins (1 and 2; *PS1* and *PS2*) cause autosomal dominant, early-onset forms of AD and account for ~1% and ~50% of inherited cases, respectively. Polymorphisms in the
10 *apoE4* and α -2 *macroglobulin* genes are associated with increased risk in individuals over 60 years of age.

The presenilins are polytopic membrane proteins expressed in the endoplasmic reticulum, Golgi complex in dendrites (close to dendritic spines) and
15 axon terminals in neurons. The *PS1* holoprotein is subject to endoproteolysis; the resulting N- and C-terminus fragments bind to each other at stoichiometric levels and/or other proteins, such as γ -catenin. The levels of the fragments are very tightly regulated and overexpression studies show little changes in the relative amounts of accumulated fragments.

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The normal biological function(s) of presenilins are not well understood although they have been shown to play a major role in the embryonic development of the axial skeleton and cerebral vasculature. The inheritance pattern in humans carrying mutant presenilin genes suggests a gain-of-function. Several cellular
25 effects of mutant presenilins have been documented that may be relevant to the pathophysiology of AD. First, in cultured cells and transgenic animals expression of

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mutant presenilins lead to the elevated production of $\alpha\beta 42(43)$ peptides that are deposited early and selectively in amyloid plaques in AD. The over-production of $A\beta 42(43)$ is most pronounced in cells expressing a PS-1 mutation lacking exon 9 ($\Delta 9$). Secondly, cells expressing mutant presenilins also have aberrant calcium homeostasis; PC12 cells expressing mutant PS1 stimulated with agonists that activate Ca^{2+} efflux from intracellular stores, exhibit larger calcium transients than cells expressing wild-type (wt) PS1.

The clinical hallmark of early AD is a disruption of memory processes. The hippocampus, which is prominently involved in the formation of memory, is affected early in the disease and shows the characteristic histopathological changes of AD, namely senile (amyloid) plaques and neurofibrillary tangles. A loss of synapses is also apparent in the hippocampus early in the disease. As the disease progresses, neuronal death in the hippocampus increases (see review by Price et al., 1998 Annu. Rev. Neurosci. 21:479-505).

Hippocampal slices have been effectively used to examine synaptic transmission and plasticity *in vitro*. Stimulation of CA1 *striatum radiatum* afferent pathways produces a mixed excitatory and inhibitory synaptic response in pyramidal neurons. Brief repetitive stimulation generates a short and long-term potentiation (STP: ~20 min and LTP: >30 min) of excitatory transmission that have been proposed as cellular correlates of some forms of learning. STP and LTP also share several underlying mechanisms with glutamate-mediated neuron death (excitotoxicity, see reviews by Obrenovitch et al., 1997 J. Prog. Neurobiol. 51:39-87 and Choi, 1992 J. Neurobiol. 9:1261-1276). All are believed to require synaptically induced postsynaptic depolarization, activation of NMDA receptors and a rise in intracellular calcium concentration. As membrane depolarization is a critical requirement for STP and LTP, these phenomena are sensitive to pharmacological manipulations of fast inhibitory pathways via the $GABA_A$ receptor. In the CA1 region of the hippocampus, for example, a $GABA_A$ antagonist leads to more LTP, while $GABA_A$ -potentiating benzodiazepines can reduce LTP. $GABA_A$ agonists can also decrease glutamate-induced excitotoxicity.

Based on the present understanding of the etiology of AD and the neuronal mechanisms associated with AD and memory, there is a need for a diagnostic method for evaluating the potential of drugs for the treatment of AD, both prophylactically and therapeutically.

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Brief description of relevant art

The effect of benzodiazepines in decreasing the incidence of AD has been described by Fastbom et al., 1998 Alzheimer Dis. Assoc. Disord. 12:14-17.

10 Mechanisms associated with learning, excitatory transmission and the involvement of GABA_A receptor are described by Bliss et al., 1993 Nature 361:31-39; Wigstrom et al., 1986 J. Physiol. (Paris) 81:228-236; Evans, et al., 1996 Neuropharmacology 35:347-357; Ohkuma et al. 1994 Jpn. J. Pharmacol. 64:125-128; and Muir et al., 1996 J. Cereb. Blood Flow Metab. 16:1211-1218. Biological functions and

15 cellular effects of presenilins are described in Shen et al., 1997 Cell 89:629-639; Wong et al., 1997 Nature 387:288-292; Lee et al., 1997 Nat. Med. 3:756-760; and Borchelt et al., 1996 Neuron 17:1005-1013. Guo et al., 1996 Neuroreport. 8:379-383 report that cells having mutant presenilins have aberrant calcium homeostasis.

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SUMMARY OF THE INVENTION

Mutant presenilin comprising hippocampal cells are employed in an assay for screening drugs for the treatment of Alzheimer's disease. Tissue samples from the hippocampus having a presenilin mutation are subjected to tetanic stimulation in the

25 presence of a candidate drug and cellular plasticity is determined, as compared to the presence of a control. The measured outcome is reduction of aberrant signaling.

BRIEF DESCRIPTION OF THE DRAWINGS

30 Fig. 1 shows graphs of a study of basal transmission and paired-pulse facilitation (PPF). Fig. 1a, top, graphs are examples of field responses in hippocampal slice CA1 region evoked by delivery of increasing intensity stimuli for wild-type ("wt")

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and $\Delta 9$ mutant animals (averages of 5 each). Fig. 1a bottom is the input-output plot of basal transmission in mutant (Δ) and wt animals (O) obtained from responses evoked as in 1a (top). The plot includes data from 4 wt and 6 mutant slices. Best fit line to each group (linear regression) shows slopes that are not significantly different ($p > 0.05$). Scale bars: 10ms and 0.2 mV. Fig. 1b top shows examples of responses to paired stimuli (50 ms inter-stimulus interval, averages of 10 each). Fig. 1b bottom is a plot of percent potentiation versus inter-pulse interval for mutant (Δ , $n=6$ slices) and wt (O, $n=6$ slices) animals. Values are not significantly different ($p > 0.05$). Scale bars: 50 ms and 0.1 mV.;

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Fig. 2 shows graphs of epsp response to tetani in the presence or absence of agents which affect the GABA_A receptor to evaluate the role of GABA_A in the presence or absence of agents which affect the GABA_A receptor. Fig. 2a graphs are based on experiments with GABA_A transmission intact. On the left, examples of epsp response (averages of 10 each) immediately before and 20 min after a tetanus superimposed for wt and mutant animals. Scale bars: 0.2 mV and 20 ms apply to a and b. On the right, plot of mean epsp slope (\pm SEM) normalized to values before tetanic stimulus (time 0). For each slice transmission two independent pathways were monitored. Tetanized pathway showed greater enhancement in slices from mutant (open triangles, $n=8$ slices) than wt (open circles, $n=10$ slices) animals. Control pathways (mutant, closed triangles; wt closed circles) remain unchanged. Tetanus consisted of 100 stimuli delivered over 1 sec (100 Hz). The potentiation at the time points of 5, 10, 15, 20, 25 and 30 min was: 1.36 ± 0.060 , 1.26 ± 0.058 , 1.27 ± 0.054 , 1.24 ± 0.058 , 1.22 ± 0.063 and 1.21 ± 0.069 , respectively for the wt. For the same time points for the mutant these values were: 1.67 ± 0.065 , 1.52 ± 0.054 , 1.54 ± 0.072 , 1.56 ± 0.075 , 1.51 ± 0.089 and 1.54 ± 0.090 , respectively. At these time points, there was a statistically significant difference between the two sets of data points (< 0.05). Abbreviation “-PTX”, no picrotoxin in the bath. Fig. 2b shows results with GABA_A transmission blocked with 100 μ M picrotoxin. On the left, are examples of epsp responses (average of 10 each) immediately before and 20 min after a tetanus superimposed for wt (top) and mutant (bottom) animals. On the right, a plot of mean epsp slope \pm SEM normalized to values before tetanic stimulus

(time 0). For these experiments, control pathways were monitored for only 30 min after tetanus. Tetanized pathways showed similar enhancement in slices from mutant (open triangle, n=13 slices) and wt (open circles, n=18 slices) animals.

Control pathways (mutant, closed triangles, wt closed circles) remain unchanged.

- 5 Tetanus consisted of 25 pulses given as groups of 5 pulses at 100 Hz every 10 s, 5 times. This tetanus was weaker than in Fig. 2a to obviate possible differences between wt and mutant induction. Abbreviation, "+PTX", with picrotoxin in bath. Fig. 2c shows the results of experiments in the presence of N-methyl-D-aspartate (NMDA)-receptor blockade with AP5. Plot of mean epsp slope \pm SEM normalized to
10 values before tetanic stimulus (time 0).;

- Fig. 3 shows the effect of flunitrazepam on LTP. Fig. 3a is a graph of the means \pm SEM of normalized epsp responses in the absence of drug plotted against time: wt (\bullet , n=7 slices), $\Delta 9$ mutation (\blacktriangle , n=12 slices). There was a significantly
15 greater amount of potentiation in the mutant at the time points of 5, 10 and 15 min post-tetanus. At 20 min, the difference in potentiation became insignificant. The tetanus (delivered at time 0) was 100 pulses given for 1s (100 Hz) every 20s 3x in succession. Control pathways (receiving no tetanus) remain unchanged. Abbreviation, "-FLU", no flunitrazepam in bath. Fig. 3b was as in Fig. 3a but in
20 the presence of flunitrazepam in the bathing medium; (\bullet , n=11 slices) and $\Delta 9$ mutation (\blacktriangle , n=8 slices). There was no statistically significant difference between the two groups at the above time points post-tetanus. Control pathways (receiving no tetanus) remained unchanged. Abbreviation, "+FLU", flunitrazepam present in bath. Fig. 3c is the same data as Figs. 3a and 3b comparing potentiation in mutants
25 (+FLU) with the wt (-FLU) to show the suppression of the potentiation to almost the wt levels. Fig. 3d are histograms from the data of Figs. 4a and 4b showing potentiation at various times post-tetanus. All groups were compared with each other, only statistically significantly different pairs ($p < 0.05$) are shown by the lines. These comparisons were calculated for 5 min, 10 min (histogram not shown) and 15
30 min. These time points gave identical statistical results for pairwise comparisons as in the 5 min case. For the 20 min time point, however, the W vs. M comparison

was not significant but the other two pairwise group comparisons were significant at $p < 0.05$. W=wild type, M=mutant, Wf=wt +FLU and Mf=mutant +FLU.;

Fig. 4 are graphs of the effect of agents on the GABA_A receptor-mediated transmission in the mutant and wt cells using whole-cell patch-clamping. Fig. 4a graphs the evoked synaptic response (averaged up to 20 each) from whole-cell patch-clamped neurons. Outward current recorded at 0mV is completely blocked by 100 μ PTX, a GABA_A receptor antagonist. NBQX blocked some of the outward current (not shown) indicating some di-synaptic inhibition. At the holding potential of -60mV, the inward current is completely blocked with 2 μ M NBQX, the AMPA (glutamate subtype) receptor blockade. Scale bar: 25 pA and 20ms. Fig. 4b graphs the evoked synaptic responses (averaged up to 15 each) recorded at holding potentials of 0mV and -60mV with the stimulating electrode placed in stratum radiatum (top) at site 1, ~50 μ from the recording electrode (middle) at site 2, ~250 μ m from the recording electrode and (bottom) at site two with the stimulus intensity increased ~3 -fold. Scale bars: top, 50pA; middle, 100 pA; bottom, 50 pA; time scale, as in Fig. 4a. Fig. 4c, left, examples of averaged (up to 15 each) traces from patch-clamp whole-cell recordings in wt and $\Delta 9$ mutation at -60mV (glutamate currents) and at 0mV (GABA_A currents). Scale bar: 40pA and 20ms. On the right, peak amplitude of response ratios (measured at holding potentials of -60mV and 0mV, respectively) from cells in individual slices (n=9 slices each). The means \pm SEM are also shown and superimposed using the filled symbols. The ratios are significantly greater in the mutant than in the wt ($p < 0.05$, t-test).

Fig. 5 graphs the results from the effects of AP5-sensitive potentials during tetanus. Fig. 5a shows normalized traces of field potential response to four consecutive (every 10ms) stimuli, before (the larger response) and after (the intermediate response) the application of the specific NMDA-receptor antagonist, AP5. The differences between the two responses at each time point are also shown. The responses were normalized to the area up to the peak of the first response (which is mostly due to non-NMDA receptor activation). Scale bar: 10ms. Fig. 5b shows the averaged differences of areas under the four response curves, before and after AP5

application in individual (wt and $\Delta 9$ mutant) slices to show the effect of tetanus on the NMDA (or AP5)-sensitive component. The means \pm SEM of all slices are also shown in filled symbols. Although the mean AP5-sensitive potentials were smaller in the mutants (despite manifesting a greater potentiation (see Fig. 2), there was no statistically significant difference between the two groups.

DETAILED DESCRIPTION OF THE INVENTION

A method for screening drugs is provided for determining their potential for the treatment of Alzheimer's disease (AD). The effect of agents on changes in plasticity of mutant cells is related to their ability to treat AD. It is found that cells with presenilin mutations, particularly PS-1, can be used in a battery of tests to evaluate plasticity of cells to tetani, where restoring wild-type behavior indicates potential use as a therapeutic.

Mammalian species may be used as a source of mutant hippocampal tissue. Any mutant which provides the desired enhanced synaptic potentiation upon tetanic stimuli in the same manner as observed with a PS-1 mutation may be employed. This can be achieved in a variety of ways of varying convenience. A transgenic mammalian host can be employed where a mutated presenilin gene is introduced, where it acts as an autosomal dominant allele. Alternatively, one may provide a transgenic host, where presenilin antisense is transcribed from an inducible promoter. Also, one can infect cells or tissue with viruses which provide such genetic capability as described above. In some situations, transformed or otherwise immortalized hippocampal cells may be employed for genetic modification. Other techniques may also be used to provide the desired mutant. The mutation is in a presenilin gene, particularly PS-1. While any mammal may be used as the source of the tissue, for convenience murine species, rats and mice, may be employed, although primates other than humans, or domestic animals, such as porcine, feline, canine, lagomorpha, etc. may also find use. Lee et al., 1997 Nat. Med. 3:756-760, describes hyperaccumulation of FAD-linked presenilin variants in vivo.

In carrying the assay out, there may be an interest in first determining synaptic transmission and plasticity in hippocampal slices of wild-type and mutant hosts. Synaptic transmission is elicited by delivering stimuli of different intensities to afferent pathways. Input-output curves are generated by plotting the slopes of
 5 excitatory postsynaptic potentials (epsp) versus fiber volley amplitude (a measure of the number of presynaptic fibers activated). Appropriately, no significant difference should be observed between wt and mutant tissue for use in the assay.

While desirably, one may have wild-type hippocampal cells matched to the
 10 mutant cells, by having substantially no genetic difference affecting the assay, as a control, such control is not essential. By knowing the response of the wild-type cells to tetanic stimuli under the conditions of the assay, one can compare the results of the mutant cells with the known standard results. However, it will usually be desirable to have wild-type matched hippocampal cells to ensure that the observed
 15 results with the mutant have a direct comparison under the conditions of the assay. The control may be performed with and/or without the candidate drug to provide a comparison with the results from the mutant cells. In addition, one may have a comparison as to the effect of a known drug having a known activity on the mutant cells under the conditions of the assay. In this way one can directly compare the
 20 activity of the candidate drug to a known drug, as well as the activity of the candidate drug on wild-type cells in relation to the synaptic potential response to tetani. The assay is usually carried out over an extended period of time taking readings at different time points and determining the potentiation. Normally, the GABA_A transmission by the cells will be intact.

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The effect of tetanic stimulus on transmission is examined in the presence of intact GABA_A receptor-mediated inhibition. In wt animals, a tetanus produces a moderate amount of potentiation. (Fig. 2a) In mutant animals, the potentiation following tetanus is greater than in wt animals. (Fig.. 2a) Differences in
 30 potentiation between wt and mutant animals is statistically tested at various time points during a course of under 60 min post-tetanus and is found to be greater in mutants. LTP assessed in the presence of an NMDA receptor antagonist results in

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blocking potentiation in both the wt and mutant cells, showing that the mutant cells have enhanced potentiation and potentiation requires NMDA-receptor activation in both types of cells.

5 Blockade of the GABA_A receptor also differentiates the response between mutant and wt cells. While GABA_A receptor blockade increases LTP in wt animals, there is no significant increase in the mutant cells. It is concluded that the effect of the mutation on potentiation is occluded by blockade of inhibition, indicating that the two factors act on a common pathway.

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 In another test the effect of an agent on LTP with a moderately strong tetanus, e.g. 3 1sec 100 Hz tetani), the potentiation is larger in mutant as compared to wt animals. However, with agents that increase GABA_A receptor transmission, suppression of the enhanced potentiation should be observed. This can be
15 demonstrated with flunitrazepam as a control or standard with which the effect of the candidate agent may be compared.

 Finally, the ratio of peak inhibitory to excitatory responses is significantly greater in mutants as compared to wt. It appears that the observed result is relatively
20 independent of the site of stimulation of the tissue and variations in stimulus intensity. Because of the greater ratio for mutant cells as compared to wt, depending upon the pathway and component of the pathway upon which the agents acts mutants may have a greater response to agents in the reduction of the difference in ratio between mutant and wt cells. This abnormality in mutants (increased
25 inhibitory transmission) is indicated to be a homeostatic (feedback) system that has been turned on in these animals to suppress the underlying aberrant signaling (increased calcium rise). Candidate drugs may not directly affect the inhibitory transmission and still be efficacious, for example, if they act to suppress calcium rise through some other mechanism under the conditions of the assay.

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Based on the tests described above, it appears that the presenilin mutation and GABA_A receptor transmission act on the same pathway that regulates potentiation of synaptic transmission. This can be explained by the mutation decreasing GABA_A receptor transmission or the mutation acts downstream of
5 GABA_A receptor transmission, along the signal transduction pathway that generates potentiation. By measuring the effect of an agent on plasticity of mutant cells as compared to wt cells, one may influence the pathway associated with the GABA_A receptor and restore the response toward the wt response.

10 It is evident from the above results that methods are provided employing mutated mammalian hippocampal cells, conveniently as tissue, which differ from wild-type cells in their increased potentiation as evidenced in their response to tetani. Furthermore, drugs can be screened to determine their effect on returning the response of the mutated cells to a wild-type response. Particularly, a mutation in
15 presenilin protein, which enhances excitability of the cells upon stimuli, allows for screening of drugs which restore wild-type behavior, as demonstrated with a benzodiazepine. By employing tetani under conditions where plasticity of the cells can be determined, an efficient screening tool is provided for determining effectiveness of drugs for the treatment of Alzheimer's disease.

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The references described throughout this specification are fully incorporated by reference. Having now fully described the invention, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the invention as set forth herein.

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